

Engineering of the Non-Model Yeast *Issatchenkia orientalis* to Produce Organic Acids

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Project Goals: The goal of this research is to engineer a non-model yeast, *Issatchenkia orientalis*, to produce organic acids in high titers. 3-Hydroxypropionic acid is one of the key sugar-based building block chemicals that can be produced in biorefineries. The ability to produce 3-hydroxypropionic acid from glucose or other renewable carbon sources would provide a biosustainable alternative to acrylic acid production from fossil resources.

The nonconventional yeast *Issatchenkia orientalis* is a potential platform microorganism for production of organic acids thanks to its unusual ability to grow in highly acidic conditions. However, the lack of efficient genetic tools, including a stable episomal plasmid and precise genome editing tool, prevented metabolic engineering of this organism. We previously developed this genetic toolbox to efficiently engineer this non-model yeast to produce value-added chemicals.^{1,2} Here we present the production of 3-hydroxypropionic acid (3HP) from *I. orientalis*. There are multiple pathways to produce 3HP; however, only two are generally used, the malonyl Co-A pathway and the β -alanine pathway. We have generated both pathways on a plasmid to determine preliminary production quantities of 3HP in shake flasks as well as the dependence of pH on the production. Our highest production of 3HP thus far is ~1.5 g/L from 50g/L glucose as the carbon source over 5 days fermentation from the β -alanine pathway. The malonyl Co-A pathway yielded only ~700 mg/L. Further experiments will be done utilizing the β -alanine pathway to further improve production by improving the expression of the beta alanine pyruvate amino transferase as well as genome engineering of the base strain.

References

1. Tran, V., Cao, M., Fatma, Z., Song, X., Zhao, H. *mSphere*, 2019, e00345-19.
2. Cao, M., Fatma, Z., Song, X., Hsieh, P.H., Tran, V., Lyon, W.L., Sayadi, M., Shao, Z., Yoshikuni, Y., Zhao, H. *Metabolic Engineering*, 2020, 59:87-97.

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